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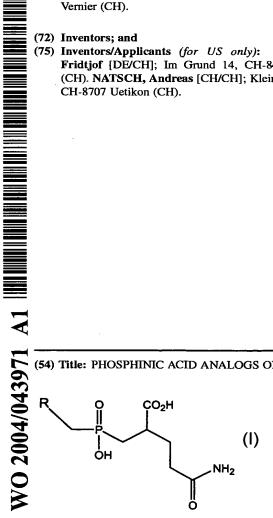
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(54) Title: PHOSPHINIC ACID ANALOGS OF GLUTAMATE



(57) Abstract: Inhibitors of axillary malodour having the formula (I): wherein R has the same meaning as given in the specification.

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PHOSPHINIC ACID ANALOGS OF GLUTAMATE

This invention is concerned with compounds useful for the prevention or suppression of human malodour, in particular human axillary malodour.

It is known that fresh sweat is odourless and that odour is only formed upon contact of sweat with skin bacteria (for example bacteria of the genera of Staphylococcus and Corynebacteria) and it is believed that odourless molecules present in sweat are degraded by bacteria colonising the axilla. It is generally accepted (Labows et. al., Cosmet. Sci Technol. Ser. (1999), 20:59-82) that highly unpleasant malodour is released from fresh sweat mainly by the Corynebacteria genus of bacteria. The principal constituents thought to be responsible for malodour include volatile steroids, volatile sulphur compounds and short-chain, branched fatty acids.

It has been suggested to treat malodour by eradicating the bacteria responsible for causing the odour. Indeed, commercially available cosmetic deodorants often contain antibacterial compounds that generally inhibit the growth of skin microflora.

Antibacterial compounds currently used in deodorant products include, for example Triclosan (2,4,4'-trichloro-2'hydroxy-diphenyl-ether). However, a draw-back to the use of antibacterials is the potential for disturbing the equilibrium of the skin's natural microflora.

Fatty acids, in particular short chain, branched fatty acids are known to play a role in axillary malodour, and are particularly foul smelling components of stale sweat. In copending application PCT/CH02/00262 the applicant has disclosed an enzyme that mediates in a process of transforming odourless compounds found in sweat into these malodorous fatty acids. In this co-pending application there is also disclosed a broad class of compounds having activity as inhibitors of the enzyme.

Nevertheless there remains the need to find further compounds displaying good inhibitory properties with respect to the above mentioned enzyme.

Accordingly, the invention provided in a first aspect a compound of formula (I)

wherein R is a substituted alkyl, benzyl or allyl residue selected from the group consisting of

- a) nonyl;
- b) 4,4,4-trifluoro-propyl;
- c) 2-methyl-4-phenyl-butyl;
- d) 4-trifluoromethyl-phenyl;
- e) pentafluorophenyl;
- f) 4-fluoro-phenyl;
- g) naphthalene-2-yl;
- h) biphenyl-2-yl;
- i) 5,5,7,8,8-pentamethyl-5,6,7,8-tetrahydro-naphthalene-2-yl;
- k) 5,5,8,8-tetramethyl-5,6,7,8-tetrahydro-naphthalen-2-yl;
- 1) 1,1,3,3-tetramethyl-indan-5-yl;
- m) styryl;
- n) 2,6-dimethyl-heptyl;
- o) 2-(4-tert-Butyl-phenyl)-1-methyl-vinyl;
- p) 2-(4-Isopropyl-phenyl)-1-methyl-vinyl;
- q) 1-(1,7,7-Trimethyl-bicyclo[2.2.1]hept-2-yl)-ethyl;
- r) 2-(4-Isobutyl-phenyl)-1-methyl-vinyl;
- s) 2-(2-isopropyl-phenyl)-1-methyl-ethenyl;
- t) 2-phenyl-ethyl;
- u) cyclohexyl-methyl;
- v) 2,2-dimethyl-propyl;
- w) 2-(pentafluorophenyl)-ethyl;
- x) 3-phenyl-propyl;
- y) heptyl;
- z) 4-isopropyl-cyclohex-1-enyl;
- za) decyl;

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- zb) hexyl;
- zc) trans-4-isopropyl-cyclohexyl;
- zd) 5-ethyl-2-methyl-heptyl;
- ze) 2,6,10-trimethyl-undecyl;
- zf) 1-methyl-3-(2,2,3-trimethyl-cyclopentyl)-propyl; and
- zg) octyl.

Compounds of the formula (I) contain chiral atoms and as such they can exist as isomeric mixtures or they may exist as pure stereoisomers. Most preferred are compounds have an S-configuration on the carbon atom in the position alpha to the carboxyl group.

As stated hereinabove, compounds of the present invention may interact with an enzyme thereby to reduce the enzyme's ability to cleave compounds in sweat leading to release of malodorous acids from odourless fresh sweat. That enzyme, described in the afore-mentioned co-pending application, was isolated from the bacteria of the genus *Corynebacteria* that can be found colonising the axilla, in particular certain *Corynebacteria sp.*, more particularly *Corynebacterium striatum* Ax 20 which has been submitted on the 26, April 2001 to the International Depository Authority DSMZ-Deutsche Sammlung von Mikrooganismen und Zellkulturen GmbH, D-38124 Braunschweig. The Accession Number provided by the International Depository Authority is DSM 14267. The enzyme was found to occur intracellularly and can be released from the cells by mechanical disruption of the cell envelope. Thus, it may be isolated from cellular extracts obtained from wild-type bacterial strains, especially from strains of Corynebacteria isolated from the human axilla, in particular Corynebacterium striatum Ax 20. In the alternative, it may be produced by recombinant means which are well known to persons skilled in the art.

The amino acid sequence of this enzyme is set forth in SEQ ID No. 1 and a nucleic acid sequence encoding for this enzyme is set forth in SEQ ID No. 2, both of which sequences are shown below.

Compounds of the present invention display inhibition of the enzyme at concentrations of about 1 to 500,nanomolar more particularly from 5 nanomolar 500 nanomolar concentration in vitro, e.g. from 9 to 150 nanomolar. Furthermore, having regard to the lipophilicity of the residue R, the compounds are adapted to penetrate the cell walls of the enzyme-producing bacteria, as such, they are efficaceous in vivo.

Indeed, the nature of the residue R appears to influence the ability of compounds to penetrate the cellular walls of different bacteria colonising the axilla and which are implicated in malodour production. For example, other strains of Corynebacteria, for example Corynebacterium bovis and Corynebacterium jeikeium, or bacteria of the genus Staphylococci found in the microflora of the axilla also produce related enzymes that themselves mediate in biochemical reactions wherein L-glutamine derivatives are cleaved at N_{α} . The compounds of the present invention may interfere in cellular processes of a wide variety of bacterial strains thereby resulting in the suppression or prevention of malodour from these sources.

The in vitro activity of the compounds as inhibitors may be measured in terms of either their IC_{50} values or their Ki values, both of which measures are well known to the person skilled in the art. As is well known, the IC_{50} value provides the concentration of an inhibitor needed to reduce enzyme velocity by half at a given substrate concentration. This value is dependent on the affinity of the substrate for the enzyme which is reflected in the value K_m of the substrate. In this way, the Ki value may be determined for a given substrate and a given substrate concentration by measuring IC_{50} and then calculating according to the following formula

$$K_{l} = \frac{IC_{50}}{1 + \frac{[Substrate]}{K_{m}}}$$

The uptake of the compounds in bacterial cells and the inhibition of the enzyme contained therein may be measured using an assay based on stationary-phase living cells. Thus, cells may be incubated along with inhibitory compound or compounds, and

the substrate (i.e. the material found in sweat, which when cleaved by the enzyme forms the malodorous acids), and the release of acids may be measured at various inhibitor concentrations. By comparing IC_{50} values obtained with the living cells with the IC_{50} values obtained with the isolated enzyme, the ease of penetration of the compounds into the bacterial cells can be assessed.

Compounds of the present invention may be added to any cosmetic and personal care products such as sticks, roll-ons, pump-sprays, aerosols, deodorant soaps, powders, solutions, gels, creams, sticks, balms and lotions to enhance the deodorising effect of these products. Preferably, a compound of the present invention may be employed in said products in amounts of about 0.01 to 0.5% by weight.

The above-mentioned products, in addition to the inhibitors, may comprise anti-bacterial agents known in the art, e.g. Triclosan. The products may also comprise dermatologically acceptable ingredients such as are commonly used in these types of product. Examples of such additional ingredients include fragrances, colorants, opacifiers, buffers, antioxidants, vitamins, emulsifiers, UV absorbers, silicones and the like. As is also well known, all products can be buffered to the desired pH.

In addition to the inhibitor, a deodorant cologne may comprise ethanol and fragrance. Fragrance may be present from 1 to 10% and the ethanol can be present to make up the mass to 100%.

Additional ingredients in a typical ethanol-free deodorant stick may include polyols, such as propylene glycol; derivatives thereof, such as propylene-glycol-3-myristyl ether (Witconol APM); water; a surfactant such as sodium stearate; and fragrance. The polyol may be present in an amount of 30 to 40%; the derivatives of the polyol likewise may be present at about 30 to 40%; water may be present to about 10 to 20%; the surfactant may be present to 5 to 10%; and the fragrance may be present in an amount up to 10%.

A typical antiperspirant stick might contain as additional ingredients such as Ethylene Glycol Monostearate (e.g. from 5 to 10%); Shea butter (e.g. from 3 to 5%); Neobee

1053 (PVO International) (e.g. from about 12 to 15%); Generol 122 (Henkel) (e.g. from about 3 to 7%); Dimethicone (DC 345)(e.g. from 30 to 40%); aluminium sesquichlorohydrate (e.g. from about 15 to 20%); and a fragrance, e.g. from 1 to 10%.

An antiperspirant aerosol may contain ethanol, e.g. from about 10 to 15%; zirconium aluminium tetrachlorohydrate, e.g. from about 3 to 5%; Bentone 38, e.g. from about 1 to 2%; fragrance in an amount aforementioned; and a hydrocarbon propellant, e.g. S-31, up to 100% based on the total aerosol composition.

An antiperspirant pump composition may contain aluminium sesquichlorohydrate, e.g. from 15 to 25%; water, e.g. from 50 to 60%; Triton X-102 (Union carbide), e.g. from 1 to 3%; dimethyl Isosorbide (ICI), e.g. from 15 to 25 %; and a fragrance in an amount as aforementioned.

All percentages mentioned above are in wt %.

Accordingly, the present invention relates to the use of the compounds of formula (I) and/or compositions containing same for the elimination or suppression of malodour. The invention also relates to compositions comprising an odour-suppressing quantity of an inhibitor compound, which acts as an inhibitor of the enzyme, and dermatologically acceptable vehicles that are generally well known in the art of cosmetic and personal care products.

The invention also provides in another of its aspects, a method of suppressing axillary malodour comprising the step of providing a composition for application to the skin of a person in need of treatment, said composition containing an inhibitor compound and dermatologically acceptable vehicle therefor, said compound being selected from one or more compounds of formula (I) described above.

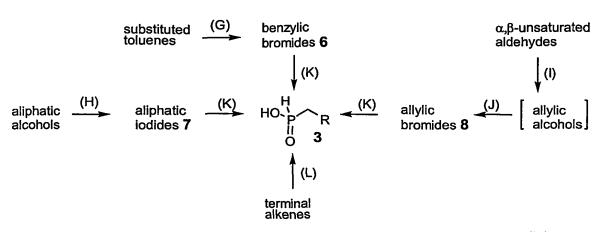
A compound of formula (I) may be prepared according to synthetic protocols as set out in detail below with reference to Scheme 1, Scheme 2 and the Examples

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- A) 5 equivalents (eq.) HP(OTMS)₂ for 2 h at 130°C.
- B) 1 eq benzylic or allylic bromide (6 or 8), 3 eq BSA, at 25°C. The product (4) is obtained in quantitative yield.
- C) 1 eq alkylphosphinic acid (3), 5 eq HMDS for 3 h at 130°C, then 1 eq acrylate (1) for 4 h at 130°C, then EtOH at 70°C. The product (4) is obtained in quantitative yield.
- D) 10-20 weight-% Pt/C, 1 atm H₂, AcOEt / EtOH 2:1, 25°C. Or Raney-Ni, EtOH, 25°C, 1 atm H₂.
- E) 1 N LiOH / EtOH for 1 day at 25°C to give the product in quantitative yield.
- F) 2.2 eq (iPr)₃SiH in TFA at 25°C for 3 h to provide compounds of the present invention.

The acrylate starting material (1) may be formed according to a method described in copending application PCT/CH02/00262.

The alkyl, benzyl or allyl halides (7,6,8) are either commercially available or may be formed from commonly available starting materials according to synthetic protocols known per se and set out in Scheme 2 below.



Scheme 2

- G) 1 eq Br₂ at 170°C for 4 h.
- H) 2 eq Pyridine, 1.2 eq PPh₃, 1.2 eq Iodine at 0°C for 2 h.
- I) 0.35 eq NaBH₄, MeOH for 2 h at 0°C, to provide the allylic alcohols in quantitative yield.
- J) Et₂O, 0.4 eq PBr₃ for 5 h at 0°C to provide the allylic bromides in quantitative yields.
- K) 3-5 eq HP(OTMS)₂ in CH₂Cl₂, for 16 h.
- L) 2 eq NaH₂PO₂(H₂O), 1 eq BEt₃, MeOH for 6 h at 25°C.

There now follows a series of Examples that serve to illustrate the invention.

Examples

The following compounds are formed according to the following syntheses:

- 5a 4-Carbamoyl-2-(decyl-hydroxy-phosphinoylmethyl)-butyric acid
- 5b 4-Carbamoyl-2-[hydroxy-(4,4,4-trifluoro-butyl)-phosphinoylmethyl]-butyric acid
- **5c** 4-Carbamoyl-2-[hydroxy-(3-methyl-5-phenyl-pentyl)-phosphinoylmethyl]-butyric acid
- **5d** 4-Carbamoyl-2-[hydroxy-(4-trifluoromethyl-benzyl)-phosphinoylmethyl]-butyric acid
- **5e** 4-Carbamoyl-2-(hydroxy-pentafluorophenylmethyl-phosphinoylmethyl)-butyric acid
- 5f 4-Carbamoyl-2-[(4-fluoro-benzyl)-hydroxy-phosphinoylmethyl]-butyric acid

- 5g 4-Carbamoyl-2-(hydroxy-naphthalen-2-ylmethyl-phosphinoylmethyl)-butyric acid
- 5h 2-(Biphenyl-2-ylmethyl-hydroxy-phosphinoylmethyl)-4-carbamoyl-butyric acid
- 5i 4-Carbamoyl-2-[hydroxy-(5,5,7,8,8-pentamethyl-5,6,7,8-tetrahydro-naphthalen-2-ylmethyl)-phosphinoylmethyl]-butyric acid
- 5k 4-Carbamoyl-2-[hydroxy-(5,5,8,8-tetramethyl-5,6,7,8-tetrahydro-naphthalen-2-ylmethyl)-phosphinoylmethyl]-butyric acid
- 5l 4-Carbamoyl-2-[hydroxy-(1,1,3,3-tetramethyl-indan-5-ylmethyl)-phosphinoylmethyl]-butyric acid
- 5m E-4-Carbamoyl-2-[hydroxy-(3-phenyl-allyl)-phosphinoylmethyl]-butyric acid
- 5n 4-Carbamoyl-2-[(3,7-dimethyl-octyl)-hydroxy-phosphinoylmethyl]-butyric acid
- **50** E-2-{[3-(4-tert-Butyl-phenyl)-2-methyl-allyl]-hydroxy-phosphinoylmethyl}-4-carbamoyl-butyric acid
- **5p** E-4-Carbamoyl-2-{hydroxy-[3-(4-isopropyl-phenyl)-2-methyl-allyl]-phosphinoylmethyl}-butyric acid
- **5q** 4-Carbamoyl-2-{hydroxy-[2-(1,7,7-trimethyl-bicyclo[2.2.1]hept-2-yl)-propyl]-phosphinoylmethyl}-butyric acid
- 5r E-4-Carbamoyl-2-{hydroxy-[3-(4-isobutyl-phenyl)-2-methyl-allyl]-phosphinoylmethyl}-butyric acid
- 5s E-4-Carbamoyl-2-{hydroxy-[3-(2-isopropyl-phenyl)-2-methyl-allyl]-phosphinoylmethyl}-butyric acid
- 5t 4-Carbamoyl-2-[hydroxy-(3-phenyl-propyl)-phosphinoylmethyl]-butyric acid
- 5u 4-Carbamoyl-2-[(2-cyclohexyl-ethyl)-hydroxy-phosphinoylmethyl]-butyric acid
- 5v 4-Carbamoyl-2-[(3,3-dimethyl-butyl)-hydroxy-phosphinoylmethyl]-butyric acid
- 5w 4-Carbamoyl-2-[hydroxy-(2-pentafluorophenyl-ethyl)-phosphinoylmethyl]-butyric acid
- 5x 4-Carbamoyl-2-[hydroxy-(4-phenyl-butyl)-phosphinoylmethyl]-butyric acid
- 5y 4-Carbamoyl-2-(hydroxy-octyl-phosphinoylmethyl)-butyric acid
- 5z 4-Carbamoyl-2-[hydroxy-(4-isopropyl-cyclohex-1-enylmethyl)-phosphinoylmethyl]-butyric acid
- 5za 4-Carbamoyl-2-(hydroxy-undecyl-phosphinoylmethyl)-butyric acid
- 5zb 4-Carbamoyl-2-(heptyl-hydroxy-phosphinoylmethyl)-butyric acid

5zc 4-Carbamoyl-2-[hydroxy-(4-isopropyl-cyclohexylmethyl)-phosphinoylmethyl]-butyric acid

5zd 4-Carbamoyl-2-[(6-ethyl-3-methyl-octyl)-hydroxy-phosphinoylmethyl]-butyric acid

5ze 4-Carbamoyl-2-[hydroxy-(3,7,11-trimethyl-dodecyl)-phosphinoylmethyl]-butyric acid

5zf 4-Carbamoyl-2-{[hydroxy-[2-methyl-4-(2,2,3-trimethyl-cyclopentyl)-butyl]-phosphinoylmethyl}-butyric acid

5zg 4-Carbamoyl-2-(hydroxyl-nonyl-phosphinoylmethyl)-butyric acid.

Structures of these compounds are set out below:

The following Examples are described with reference to Scheme 1 and Scheme 2. All compounds referred to in the Examples are defined by the combination of the corresponding compound number given in Scheme 1 or Scheme 2 and the letter code of the corresponding "R" residue. For example (41) stands for compound 4 of Scheme 1, wherein R is 1,1,3,3-tetramethyl-indan-5-yl.

Example 1:

A) Preparation of 2-Hydroxyphosphinoylmethyl-4-(trityl-carbamoyl)-butyric acid ethyl ester (2) (Step A of Scheme 1)

In a 500 mL flask equipped with a septum and a condenser, 25 g (0.3 mol) ammonium phosphinate and 49 g (0.3 mol) HMDS are heated under N₂ at 110°C for 3.5 h. The reaction mixture is cooled to 5°C where 25 g acrylate 1 in 150 ml dichloromethane is added. The mixture is stirred for 16 h at room temperature. Work-up: 1 N HCl and CH₂Cl₂ are added. The organic phase is washed with 1 N HCl, the combined acidic phases are re-extracted with CH₂Cl₂. The combined organic phases are dried over MgSO₄, evaporated under reduced pressure and dried at 50°C under high vacuum yielding 28.8 g of phosphinic acid 2.

Yield: Quant; M.p.:152-154°C (white solid); Purity: 89 % (31P-NMR)

³¹P-NMR (CDCl₃, 400MHz): 32.0 ppm (s).

MS (ESI neg.): 957 [2M-H], 478 [M-H].

¹H-NMR (CDCl₃, 400MHz): 1.2 (t, 3H), 1.8 (m, 1H), 1.9 (2H), 2.05 (m, 1H), 2.25 (m, 1H), 2.7 (m, 1H), 4.1 (q, 2H), 6.3 - 7.7 (d, 1H, P-H, J = 560 Hz), 6.9 (s, 1H, NH), 7.2 (15H, trityl-H), 8.2 (1H, P-OH).

¹³C-NMR (CDCl₃, 400MHz): 14.2 (CH₃), 28.7 (d, CH₂), 30.8, 31.8 (d, P-CH₂), 34.3 (s, CH₂), 38.2 (CH), 61.2 (OCH₂), 70.5 (Ph₃C), 127.0 (3C, Trityl-CH), 127.9 (6C, Trityl-CH), 128.7 (6C, Trityl-CH), 144.6 (3C, Trityl-C), 171.0 (C=O), 174.0 (C=O).

B) Preparation of 2-[Hydroxy-(1,1,3,3-tetramethyl-indan-5-ylmethyl)-phosphinoyl-methyl]-4-(trityl-carbamoyl)-butyric acid ethyl ester (4l) (Step B of Scheme 1)

In a 100 mL flask equipped with a septum and a condenser, monoalkylphosphinic acid 2 (3 g, 6.4 mmol) is dissolved in dry CH₂Cl₂ (20 ml). 5-Bromomethyl-1,1,3,3-tetramethyl-indan 6l (1.9 g, 7 mmol) and BSA (3.9 g, 19 mmol) are added and the mixture is stirred 72 h at 25°C. Work-up: The mixture is poured on 1N HCl. The organic phase is washed with 1N HCl, the combined acidic phases are re-extracted with 1N HCl. The combined organic phases are dried over MgSO₄, evaporated under reduced pressure and dried at 50°C under high vacuum to give 4.77 g of the bisalkylated phosphinic acid 4l.

Yield: Quant; Purity: 82 % (31P-NMR)

³¹P-NMR (CDCl₃, 400MHz): 53.8 ppm (s).

MS (ESI neg.): 1329 (10% [2M - H]), 664 (100% [M - H]), 494 (30%).

¹H-NMR (CDCl₃, 400MHz): 1.2 (t, 3H), 1.3 (14H), 1.7 (m, 1H), 1.9 (d, 2H, P-CH₂), 2.1 (m, 1H), 2.25 (2H), 2.7 (m, 1H), 3.0 (d, 2H, P-CH₂), 4.1 (q, 2H, OCH₂), 6.85 (s, 1H, NH), 7.2 (15H, trityl-H), 8.4 (s, 1H, P-OH).

Whereas this synthesis is described with reference to the "R" residue relating to compound 41 above, this synthesis is carried out for the preparation of other benzylic, or allylic phosphinoyl compounds whose "R" residues correspond to the compounds 4d, 4e, 4g, 4h, 4i, 4k, 4m, 4n, 4o, 4p, 4q, 4r, 4s, 4zd, 4ze and 4zf.

The 3 eq BSA of the above procedure can be replaced by 5-7 eq HMDS and the workup can be simplified by addition of ethanol followed by concentration. In this way 4e, 4f and 4n were prepared.

C) Preparation of 4-Carbamoyl-2-[hydroxy-(4,4,4-trifluoro-butyl)-phosphinoylmethyl]-butyric acid (4b): (Step C of Scheme 1)

0.3 g (1.7 mmol) (4,4,4-trifluoro-butyl)-phosphinic acid 3b (0.3 g, 1.7 mmol) is dissolved in HMDS (1.4 g, 8.5 mmol) at room temperature and heated at 130°C for 4 h. At 80°C acrylate 1 (0.7 g, 1.7 mmol) is added and the reaction mixture heated at 130°C for 16 h. Ethanol is added at 60°C and the mixture refluxed for 30 min. The solvents are removed under reduced pressure and the residue is dried at 50°C under high vacuum for 8 h to yield 0.9 g of 4b.

Yield: 89 %; Purity: ~80 % (1H-NMR)

³¹P-NMR (CDCl₃, 400MHz): 45.3 ppm (s).

MS (ESI neg.): 670 (8% [M + NaOAc - H], 588 (100% [M - H]).

¹H-NMR (CDCl₃, 400MHz): 1.2 (t, 3H), 1.55 (m, 2H), 1.7 (m, 1H), 1.8 (3H), 1.95 (m, 1H), 2.15 (3H), 2.2 (m, 1H), 2.4 (m, 1H), 2.75 (m, 1H), 4.1 (q, 2H), 6.8 (s, 1H, NH), 7.2 (Trityl-H).

Whereas this synthesis is described with reference to the "R" residue relating to compound 4b (above), this synthesis is carried out for the preparation of other phosphinoyl compounds whose "R" residues correspond to the compounds 4a, 4c, 4f, 4i, 4t, 4u, 4v, 4w, 4x, 4y, 4z, 4za to 4zc and 4zg.

Example 2: Preparation of 4-Carbamoyl-2-[(3,7-dimethyl-octyl)-hydroxy-phosphinoyl-methyl]-butyric acid (5n) (Steps D,E,F of Scheme 1):

Step D: 288 g (0.4 mol) of the P-geranyl phosphinoyl compound 4n (prepared according to Example 1B) is dissolved in 1.4L ethanol at 70°C. 58 g of platinum (2.5% on charcoal / H₂O 1:1) is added at room temperature and the mixture is stirred under hydrogen for several days until complete hydrogenation (of the 2 double bonds) is detected by ¹H-NMR or MS / ESI. The mixture is filtered over Celite which is washed with 0.3L ethanol.

Step E: 2L LiOH (1N in H_2O) are added to the filtrate. Under stirring the solution is heated to 50°C for 1-2 days until complete hydrolysis is detected by ¹H-NMR or MS / ESI. The solution is neutralized by addition of ca. 250 ml conc. HCl. The supernatant solution is decanted from the precipitates, the ethanol is removed from this solution

under reduced pressure and the remaining H₂O phase is extracted with 3 x 0.5L CH₂Cl₂. The above precipitates are dissolved in 1.5L CH₂Cl₂. The combined organic layers are dried over MgSO₄, filtered and concentrated under reduced pressure to give 206 g of a reddish solid.

Step F: The resulting material of Step E (ca. 0.35 mol) is dissolved in 2.5L trifluoroacetic acid. 120 g (0.75 mol) triisopropylsilane is added and the resulting suspension stirred for 4 h at room temperature. The trifluoroacetic acid is removed under reduced pressure. 3L H₂O is added to the residue und the resulting suspension stirred at 60°C for 15 min. The supernatant water-phase is decanted off. 3L NH₃ (6% in water) is added to the residue, the resulting suspension is stirred at 60°C for 15 min and filtered over Celite. The filtrate is divided into 3 portions and filtered over 3 Chromabond C18-columns filled each with 10 g RP (Reverse Phase) material. The filtrates are concentrated under reduced pressure. To the residues toluene is added and removed under reduced pressure (3 times). Drying under high vacuum gives 109 g of a white-yellow solid foam.

Yield: 78% (based on substrate 4n); Purity: 90 % (³¹P-NMR)

³¹P-NMR (D₂O, 400MHz): 44.8 ppm (s).

MS (ESI neg.): 348 (100% [M-H]).

¹H-NMR (D₂O, 400MHz): 1.2 (2d, 9H), 1.0-1.7 (13H), 1.8-2.0 (3H), 2.2-2.3 (t, 2H, P-CH₂), 2.5-2.6 (1H), 4.8 (4H, CO₂H, POH, NH₂) ppm.

¹³C-NMR (D₂O, 400MHz): 18.7, 18.8, 22.4, 22.5, 24.6 (CH₂), 27.7, 27.2 and 28 (d, P-CH₂), 29.3 (CH₂), 30.7 and 31.7 (d, P-CH₂), 32.8 (CH₂), 33,7, 33.8, 36.5 (CH₂), 39.1 (CH₂), 40.0, 178.2 (C=O), 179.6 (C=O).

Whereas this synthesis is described with reference to the "R" residue relating to compound 5n (above), this synthesis is carried out for the preparation of other phosphinoyl compounds whose "R" residues correspond to the compounds 5zc, 5zd, 5ze and 5zf which are derived from P-alkyl-, and P-γ,γ-disubstituted allyl-phosphinoyl compounds 4zc, 4zd, 4ze, and 4zf which are prone to oxaphospholane formation under acidic conditions. Therefore the corresponding C=C-double bonds were removed by hydrogenation, prior to hydrolysis / detritylation.

All other phosphinoyl compounds (5a – 5m, 50 to 5zb and 5zg) were prepared without prior hydrogenation (Step D) just by hydrolysis / detritylation (Steps E and F). Compound 5zc was also prepared following the general procedure of Example 2 except that the hydrogenation was carried out over Raney-Nickel in place of platinum on charcoal (Step D).

Example 3: Preparation of 7-Bromomethyl-1,1,2,4,4-pentamethyl-1,2,3,4-tetrahydronaphthalene 6i: (Step G of Scheme 2)

In a 100mL flask equipped with thermometer, septum and a condenser 21.6 g (0.1 mol) 1,1,2,4,4,7-hexamethyl-1,2,3,4-tetrahydro-naphthalene (prepared as described by Wood, T. F.; Easter, W. M., Jr.; Carpenter, M. S.; Angiolini, J. Org. Chem.28, 2248 (1963)) is heated to 170°C. 16 g (0.1 mol) Bromine is added and the reaction mixture stirred for 5 h at 170°C. The content of the flask is fractionated over a Vigreux column (110°C, 4 mbar) giving 17 g (58%) 6i as a colorless liquid.

Yield: 58%; GC-purity: 71%

GC/MS: $294 / 296 (5\%, [M]^{+})$, $279 / 281 (10\%, [M - CH_{3}]^{+})$, $215 (100\%, [M - Br]^{+})$, $201 (65\%, [M - CH_{2}Br]^{+})$, 157 (55%).

¹H-NMR (CDCl₃, 400MHz): 0.95 (d, 3H), 1.05 (s, 3H), 1.25 (s, 3H), 1.3 (s, 3H), 1.35 (s, 3H), 1.4 (dd, 1H, A), 1.65 (dd, 1H, B), 1.85 (m, 1H, CH), 4.5 (s, 2H, CH₂Br), 7.15 (d, 1H, Ar-H), 7.25 (d, 1H, Ar-H), 7.35 (s, 1H, Ar-H).

¹³C-NMR (CDCl₃, 400MHz): 16.8 (CH₃), 25.0 (CH₃), 28.5 (CH₃), 31.9 (CH₃), 32.3 (CH₃), 34.3 (CH₂), 34.4 (C), 34.5 (CH), 37.8 (C), 43.5 (CH₂Br), 126.2, 127.0, 127.7 (Ar-CH), 134.7, 145.2, 146.5 (Ar-C).

Example 4: Preparation of (5-Iodo-3-methyl-pentyl)-benzene 7c: (Step H of Scheme 2) 5 g (28 mmol) Phenoxanol (3-Methyl-5-phenyl-pentan-1-ol) is dissolved in 200 ml dichloromethane under nitrogen and stirring. Triphenylphosphine (8.8 g, 34 mmol) and 4.2 g (53 mmol) pyridine are added at 25°C. After cooling to 0°C iodine (8.5 g, 34 mmol) is added. After 2 h stirring at 0°C the reaction mixture is poured on ice-cooled 1 N HCl and extracted with dichloromethane. The combined organic phases are washed

with 10% Na₂S₂O₃, sat. NaHCO₃ and sat. NaCl. Drying over MgSO₄ and evaporation gives 17g of a residue which is triturated with hexane and filtered over a 5 cm Silicagel pad. The filtrate is evaporated under reduced pressure and dried under high vacuum giving 7.1 g of 7c as a colorless oil.

Yield: 88%; Purity: > 95% (GC, NMR)

GC/MS: 288 (5%, $[M]^{+}$), 161 (10%, $[M-I]^{+}$),

119 (5%), 105 (20%, [PhCH₂CH₂]⁺), 91(100%, [[PhCH₂]⁺).

¹H-NMR (CDCl₃, 400MHz): 0.95 (d, 3H, CH₃), 1.45 (m, 1H), 1.6 (3H), 1.9 (m, 1H), 2.6 (2H, PhCH₂), 3.2 (2H, CH₂I), 7.2 (5H, ArH).

¹³C-NMR (CDCl₃, 400MHz): 4.9 (CH₂), 18.7 (CH₃), 33.3 (CH₂), 33.7 (CH₃), 38.2 (CH₂), 40.9 (CH₂), 125.8, 128.3, 128.4 (ArCH), 142.5 (ArC).

Example 4: Preparation of E-1-(3-Bromo-2-methyl-propenyl)-4-tert-butyl-benzene 80: (Steps I and J of Scheme 2)

20 g (0.1 mol) E-3-(4-tert-Butyl-phenyl)-2-methyl-propenal (prepared according to US 4435585) is added to a stirred solution of 1.2 g (32 mmol) of sodium borohydride in 20 ml of methanol at 0°C. After 2 h at room temperature quantitative conversion is checked by TLC. The reaction mixture is poured onto 40 ml saturated sodium chloride and extracted with tert-butyl methyl ether. Drying over magnesium sulfate and evaporation of the solvent gives the crude allylic alcohol (19.2 g, 94%), which is transferred to the following bromination reaction without further purification.

5 g (24 mmol) of the crude allylic alcohol is dissolved in 35 ml of dry diethyl ether under nitrogen. At 0°C phosphorus tribromide (0.95 ml, 10 mmol) is added via syringe. The reaction is stirred at 0°C for 4 h, poured onto ice and extracted three times with diethyl ether. The organic layer is washed with saturated NaHCO₃ and dried over MgSO₄. The solvent is removed under reduced pressure giving the crude allylic bromide 5.7 g of 80.

Yield: 5.7 g (87% from the aldehyde); Purity: > 95% (GC, NMR) GC/MS: 266 / 268 (3%, [M]⁺), $251 (1\%, [M-CH_3]^+)$, $188 (25\%, [M-Br]^+)$, $173 (55\%, [M-Br-CH_3]^+)$, 157 (10%), 131 (55%), 115 (22%), 91 (16%), 57 (100%).

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¹H-NMR (CDCl₃, 400MHz): 1.3 (s, 9H, tBu-CH₃), 2.0 (s, 3H, CH₃), 4.15 (s, 2H, CH₂Br), 6.6 (s, 1H, =CH), 7.22 (d, 2H, Ar-H), 7.35 (s, 2H, Ar-H).

¹³C-NMR (CDCl₃, 400MHz): 16.6 (CH₃), 31.3 (3C, tBu-CH₃), 34.6 (tBu-C), 42.6 (CH₂), 125.2, 128.7 (ArCH), 130.0 (=CH), 133.7, 134.0 (Ar-C), 150.2 (=C)

Example 6: Preparation of (3,7-Dimethyl-octa-2,6-dienyl)-phosphinic acid 3n: (Step K of Scheme 2)

In a 750mL flask equipped with septum, thermometer and condenser, ammonium phosphinate (25 g, 0.3 mol) and HMDS (51 g, 0.32 mmol) are heated under N₂ at 110°C for 3 h. The reaction mixture is cooled to 0°C. 300 mL dried CH₂Cl₂ are added followed by the addition of geranyl bromide (13.1 g, 60 mmol). The mixture is stirred for 16 h at room temperature. 10 ml methanol are added and the fine suspension is filtered over a double filter layer. The filtrate is concentrated under reduced pressure. 10% Na₂CO₃ and *tert*-butyl methyl ether are added, the phases are separated and the alkaline layer purified with *tert*-butyl methyl ether. The alkaline layer is treated with conc. HCl until pH = 1 and is then 3 times extracted with dichloromethane. Drying of the dichloromethane layer over MgSO₄ and evaporation give 13.7 g (81%) of 3n as orange oil.

Yield: 13.7 g (81%); Purity: 77% (31P-NMR)

MS (ESI neg.): $403 (10\% [2M - H]^{+})$, $201 (100\%, [M-H]^{+})$.

¹H-NMR (CDCl₃, 400MHz): 1.2 (d, 1H), 1.6 (s, 3H, CH₃), 1.6 (6H, 2 CH₃), 2.1 (4H, CH₂CH₂), 2.6 (dd, 2H, P-CH₂), 5.1 (1H, =CH), 5.15 (1H, =CH), 6.22 and 7.6 (d, 1H, J = 548 Hz, P-H), 11.8 (s, 1H, POH).

¹³C-NMR (CDCl₃, 400MHz): 16.5 (CH₃), 17.6 (CH₃), 25.6 (CH₃), 26.4 (CH₂), 30.4 and 29.5 (d, P-CH₂), 39.7 (CH₂), 110.5 (=CH), 123.7 (=CH), 131.7 (=C), 141.9 (=C).

Example 7: Preparation of (3-phenyl-propyl)-phosphinic acid 3t: (Step L of Scheme 2) To a solution of NaH₂PO₂(H₂O) (13.2 g, 0.125 mol) and allylbenzene (6.6 g, 56 mmol) in methanol (250 ml) is added triethylborane (1M in THF, 50 ml, 50 mmol) at room

³¹P-NMR (CDCl₃, 400MHz): 34.9 ppm (s).

temperature in an open 500 ml flask. The solution is stirred for 2 h at room temperature. The reaction mixture is concentrated under reduced pressure. 100 ml saturated KHSO₄ are added to the residue followed by extraction (200, 100 and 70 ml) with ethyl acetate. To the combined ethyl acetate layers are added 40 ml 10% Na_2CO_3 . Under vigorous stirring and dropwise addition of conc. NaOH the biphasic mixture is adjusted to pH = 10. The organic phase is separated and the alkaline phase adjusted to pH = 2 by addition of conc. HCl. Extraction with chloroform (3 x 100 ml), drying of the combined organic layer over MgSO₄ and evaporation gives 5.4 g of crude 3t (61%).

Yield: 61%; Purity: 84% (31P-NMR)

³¹P-NMR (CDCl₃, 400MHz): 38.2 ppm (s).

MS (ESI neg.): $265 (6\% [M-H+NaOAc]^{+})$, $183 (100\%, [M-H]^{+})$.

¹H-NMR (CDCl₃, 400MHz): 1.75 (m, 2H, CH₂), 1.9 (m, 2H, CH₂), 2.76 (t, 2H,

 $PhCH_2$), 6.4 and 7.7 (d, 1H, J = 548 Hz, P-H), 11.4 (s, 1H, POH).

Example 8: Measuring inhibitory activity

Cellular Extracts of Corynebacterium striatum Ax 20 (DSM 14267) are prepared by mechanical disruption and subsequent centrifugation.

The extract (50 μ l ml corresponding to 0.2 ml initial cell culture) is added to 50 μ l of Buffer A (Phosphate Buffer, pH 7). Various concentrations of the compounds of the present invention are added in a volume of 40 μ l, and after 10 min preincubation at 37°C, the reaction is and amended with 10 μ l of substrate (N α -lauroyl-L-glutamine, final concentration 50 μ M). The samples are incubated for 15 min and then the reaction is stopped by adding 75 μ l of Fluorescamine (2.5 mM dissolved in Acetonitrile). The fluorescence resulting from derivatization of the released glutamine with fluorescamine is determined at an excitation wavelength of 381 nm and an emission wavelength of 470 nm. By comparing the samples containing compounds of the present invention with control samples with enzyme and substrate only, the inhibition (%) is calculated. Alternatively, the same assay is made with a recombinantly formed enzyme produced with a strain containing an expression vector comprising a nucleic acid sequence

encoding for the enzyme. The results for some compounds of the present invention are listed in Table 1.

Table 1. Enzyme inhibition

Compound	IC ₅₀ value (nM)	Compound	IC ₅₀ value (nM)
5a	30.3	5x	40
5c	11.5	5z	32
5g	137.5	5у	17.9
5t	125	5n	9

In order to evaluate enzyme activity in intact cells, stationary phase living cells of Ax20 are harvested and resuspended in Buffer A to an optical density at 600 nm of 0.25. Inhibitory compounds are added at various concentrations, and after a preincubation of 15 min, the substrate (Nα-lauroyl-L-glutamine, 1 mM final concentration) is added. The samples are incubated for 1 h and then extracted with MTBE and HCl and analysed for released lauric acid using capillary GC. By comparing the samples containing compounds of the present invention with control samples with bacteria and substrate only, the inhibition (%) is calculated. By comparing the inhibitory capacity of the compounds on the isolated enzyme with the values obtained using intact cells, the relative uptake of the compounds by the cells can be assessed. From Table 2 it appears, that compounds of the present invention can cross the bacterial cell wall and cytoplasmatic membrane, and thus can have inhibitory activity in living cells.

<u>Table 2</u>. Inhibition of the enzymatic activity in living cells at the concentration of 0.2 micromolar

Compound	% inhibition of release of fatty acids by Corynebacterium Ax20 at a concentration of $0.2 \mu M$
5a	60.9
5c	70.5
5n	69.5
5у	64.9

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BUDAPEST TREATY ON THE INTERNATIONAL RECOGNITION OF THE DEPOSIT OF MICROORGANISMS FOR THE PURPOSES OF PATENT PROCEDURE

INTERNATIONAL FORM

Givaudan SA 1214 Vernier Switzerland

RECEIPT IN THE CASE OF AN ORIGINAL DEPOSIT issued pursuant to Rule 7.1 by the INTERNATIONAL DEPOSITARY AUTHORITY identified at the bottom of this page

I. IDENTIFICATION OF THE MICROORGANISM	
Identification reference given by the DEPOSITOR: Ax 20	Accession number given by the INTERNATIONAL DEPOSITARY AUTHORITY: DSM 14267
II. SCIENTIFIC DESCRIPTION AND/OR PROPOSED TAXONOMIC DES	IGNATION
The microorganism identified under L above was accompanied by: (X) a scientific description	
(X) a proposed taxonomic designation (Mark with a cross where applicable).	
III. RECEIPT AND ACCEPTANCE	
This International Depositary Authority accepts the microorganism identified (Date of the original deposit).	inder I. above, which was received by it on 2001-04-26
IV. RECEIPT OF REQUEST FOR CONVERSION	
The microorganism identified under I above was received by this Internationa and a request to convert the original deposit to a deposit under the Budapest for conversion).	l Depositary Authority on (date of original deposit) Freaty was received by it on (date of receipt of request
V. INTERNATIONAL DEPOSITARY AUTHORITY	
Name: DSMZ-DEUTSCHE SAMMLUNG VON MIKROORGANISMEN UND ZELLKULTUREN GmbH Address: Mascheroder Weg 1b D-38124 Braunschweig	Signature(s) of person(s) having the power to represent the International Depositary Authority or of authorized official(s): Date: 2001-04-30

Form DSMZ-BP/4 (sole page) 0196

Where Rule 6.4 (d) applies, such date is the date on which the status of international depositary authority was acquired.

Sequence listing part of description

SEQUENCE LISTING

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200

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Claims

1. A compound of formula (I)

wherein R is a substituted alkyl, benzyl or allyl residue selected from the group consisting of nonyl; 4,4,4-trifluoro-propyl; 2-methyl-4-phenyl-butyl; 4-trifluoromethyl-phenyl; pentafluorophenyl; 4-fluoro-phenyl; naphthalene-2-yl; biphenyl-2-yl; 5,5,7,8,8-pentamethyl-5,6,7,8-tetrahydro-naphthalene-2-yl; 5,5,8,8-tetramethyl-5,6,7,8-tetrahydro-naphthalen-2-yl; 1,1,3,3-tetramethyl-indan-5-yl; styryl; 2,6-dimethyl-heptyl; 2-(4-tert-Butyl-phenyl)-1-methyl-vinyl; 2-(4-Isopropyl-phenyl)-1-methyl-vinyl; 1-(1,7,7-Trimethyl-bicyclo[2.2.1]hept-2-yl)-ethyl; 2-(4-Isobutyl-phenyl)-1-methyl-vinyl; 2-(2-isopropyl-phenyl)-1-methyl-ethenyl; 2-phenyl-ethyl; cyclohexyl-methyl; 2,2-dimethyl-propyl; 2-(pentafluorophenyl)-ethyl; 3-phenyl-propyl; heptyl; 4-isopropyl-cyclohex-1-enyl; decyl; hexyl; trans-4-isopropyl-cyclohexyl; 5-ethyl-2-methyl-heptyl; 2,6,10-trimethyl-undecyl; 1-methyl-3-(2,2,3-trimethyl-cyclopentyl)-propyl; and octyl.

- 2. A composition comprising a body odour-suppressing quantity of a compound claimed in claim 1.
- 3. Composition according to claim 2 wherein the compound is present in amounts of about 0.01 to 0.5 % by weight.
- 4. Composition according to claim 2 or claim 3 selected from cosmetic and personal care products, in particular deo-sticks, roll-ons, pump-sprays, aerosols, deodorant soaps, powders, solutions, gels, creams, sticks, balms and lotions.

- 5. Use of a compound as defined in claim 1 or composition comprising a compound as defined in claim 1 to inhibit an enzyme in its ability to cleave compounds contained in sweat into short-chained, branched fatty acids, which enzyme is produced in bacteria of the genus *Corynebacteria*, which bacteria has been deposited at the International Depository Authority DSMZ- Deutsche Sammlung von Mikrooganismen und Zellkulturen GmbH, D-38124 Braunschweig under the Accession Number DSM 14267.
- 6. Use of a compound as defined in claim 1 or composition comprising a compound as defined in claim 1 to inhibit an enzyme in its ability to cleave compounds contained in sweat into short-chained, branched fatty acids, which enzyme is defined by an amino acid sequence set forth as SEQ ID No. 1.
- 7. A method of suppressing axillary malodour comprising the step of providing a composition for application to the skin of a person in need of treatment, said composition containing an inhibitor compound and a dermatologically acceptable vehicle therefore, said compound being selected from one or more compounds as defined by claim 1.

INTERNATIONAL SEARCH REPORT

Internation pplication No PCT (CH 03/00750

IPC 7	C07F9/30		
According to	o International Patent Classification (IPC) or to both national classifica	ation and IPC	
	SEARCHED		
Minimum do	cumentation searched (classification system followed by classification CO7F	on symbols)	
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Documentat	ion searched other than minimum documentation to the extent that s	uch documents are included in the fields s	agrahad
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C DOCUM	ENTS CONSIDERED TO BE RELEVANT		
Category °	Citation of document, with indication, where appropriate, of the rel	evant nassages	Relevant to claim No.
	The state of the s		neievani to ciann No.
P,X	WO 02/092024 A (ACUNA GONZALO ; 6	SIVAUDAN	1-4,7
	SA (CH); GFELLER HANS (CH); NATSO	CH ANDREAS	,
	(C) 21 November 2002 (2002-11-21) cited in the application		
	page 10; claims 11-13,16-18; figu	ire 2	
A	 EP 0 265 412 A (MONSANTO CO)		,
1^	27 April 1988 (1988-04-27)		1
	page 2, line 8 - line 26		
			
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Forth	ner documents are listed in the continuation of box C.	V Detect form?	<u> </u>
		χ Patent family members are listed	in annex.
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1	6 February 2004	10/03/2004	
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Information on patent family members

Internation	pplication No
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Patent document cited in search report		Publication date		Patent family member(s)	Publication date
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EP 0265412	A	27-04-1988	AT DE EP JP US	73457 T 3777325 D1 0265412 A2 63122694 A 4764620 A	15-03-1992 16-04-1992 27-04-1988 26-05-1988 16-08-1988